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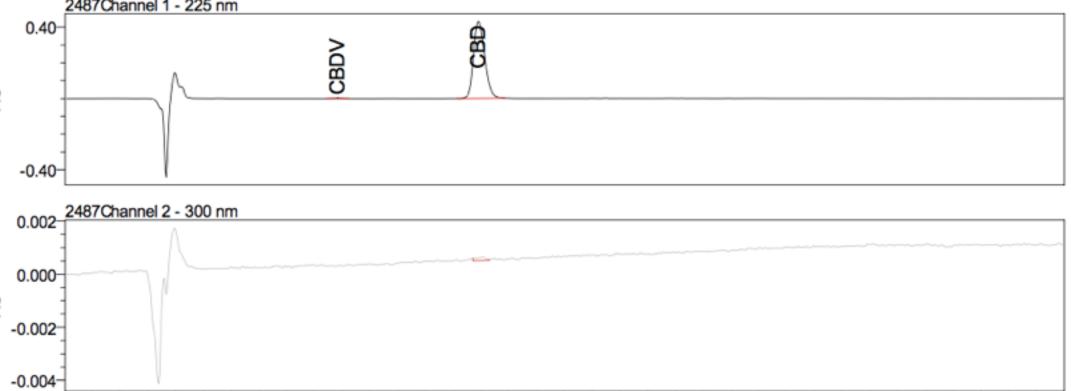
Responsible Supervisor: Sample **Date samples received:** Date analysis began: Date sample report produced: ID Number when available: Sample Mass

Martin V. **Batch# 744** <u>31-July 2017</u> 31-July 2017 01-August 2017 **1** g

Total CBD+CBDA 99.65% Cannabinoid Profile:

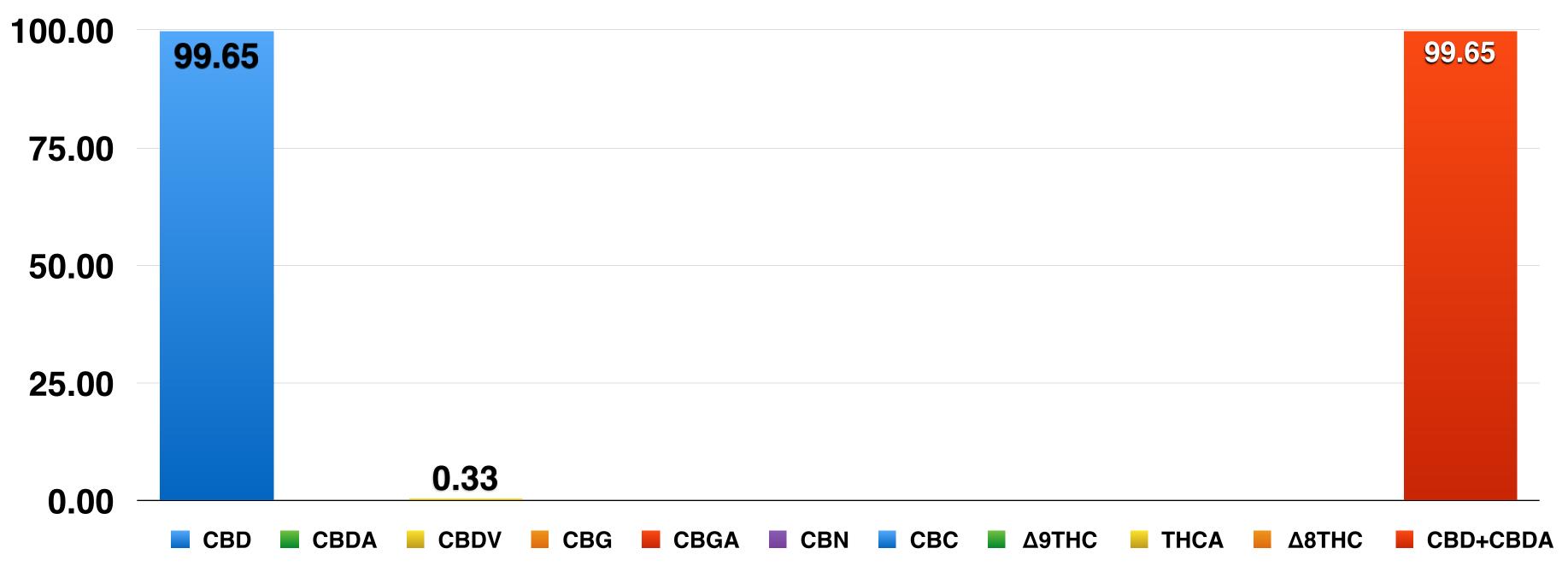
HPLC Chromatograph Raw Data

Component	Mass (%)	Amount (mg/g)	2487Channel 1 - 225 nm 0.40
CBD	99.65	996.50	A A A A A A A A A A A A A A A A A A A
CBDA	ND	ND	
CBDV	0.33	3.30	-0.40 0.002 2487Channel 2 - 300 nm
CBG	ND	ND	0.002
CBGA	ND	ND	₽ -0.002
CBN	ND	ND	
CBC	ND	ND	-0.004- 2.00 4.00 6.00 8.00 10.00 12.00 14.00 16.00 18
Δ9ΤΗϹ	ND	ND	Minutes
THCA	ND	ND	
Δ8ΤΗϹ	ND	ND	
CBD+CBDA	99.65	996.50	



ND - Not Detected

Cannabinoids as Percent of Total Mass



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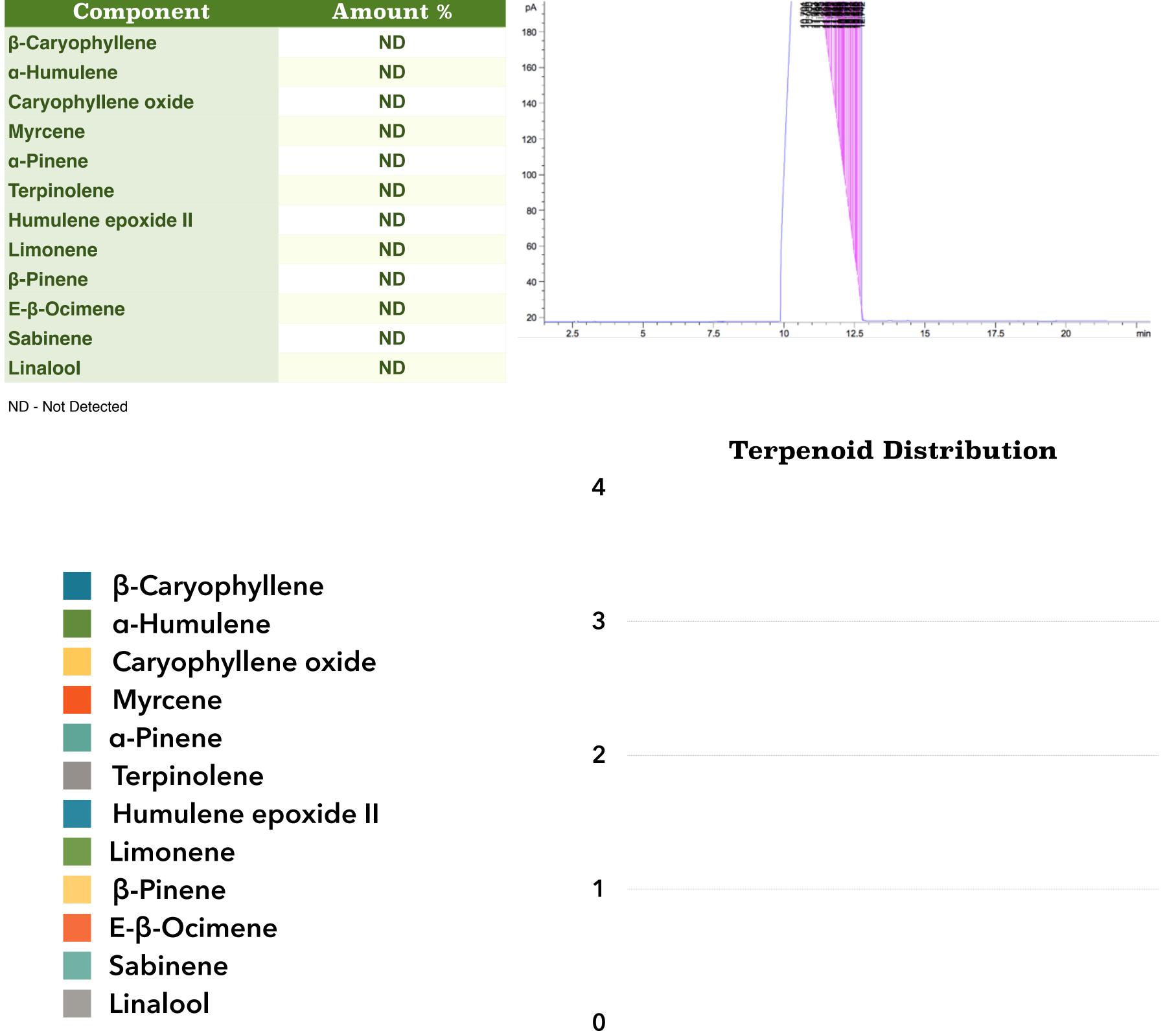
Responsible Supervisor: Sample **Date samples received:** Date analysis began: Date sample report produced: 01-August 2017 ID Number when available: Sample Mass

Martin V. **Batch# 744** 31-July 2017 31-July 2017

<u>1 g</u>

Total CBD+CBDA 99.65% Terpenoid Profile:

Component	Amount %	pА
β-Caryophyllene	ND	180 -
a-Humulene	ND	160 -
Caryophyllene oxide	ND	140 -
Myrcene	ND	120 -
a-Pinene	ND	100 -
Terpinolene	ND	100-
Humulene epoxide II	ND	80 -
Limonene	ND	60 -
β-Pinene	ND	40 -



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Responsible Supervisor: Sample **Date samples received:** Date analysis began: Date sample report produced: _____ ID Number when available: **Sample Mass**

Martin V. **Batch# 744** 31-July 2017 31-July 2017 01-August 2017 **1** g

Total CBD+CBDA 99.65% Microbial Profile:

Component		Results
Listeria m.	1 g	ND*
Escherichia c.	1 g	ND*
Salmonella	25 g	ND*
Yeast	1 g	ND*
Mould	1 g	ND*

All Mycotoxins at **Non Detectable (ND) levels**

NO MYCOTOXINS RESIDUES TO DISPLAY

G1

G2

B2

B1

Α

5

4

3

2

0

Amount (%) Mass of Mycotoxins

Nutrition Facts

Component	%
Moisture	<0.1
Protein	ND*
Total fat	ND*
Total Carbohydrates	ND*
Dietary Fibers	ND*
Sugars	ND*
Ash	ND*

*ND - Not detected

*ND - Not detected

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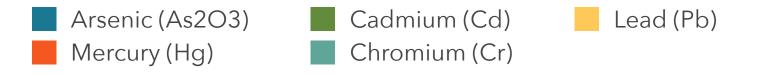
Martin V. Batch# 744 31-July 2017 31-July 2017 01-August 2017 1 g

Total CBD+CBDA 99.65% Heavy Metals Profile:

All Heavy Metals at Non Detectable (ND) levels

Component	Mass (%)	Amount (ppm)	Limit ^{**} (ppm)
Arsenic (As ₂ O ₃)	ND*	< 0.1	< 0.1
Cadmium (Cd)	ND*	< 0.1	< 0.1
Lead (Pb)	ND*	< 0.1	< 0.1
Mercury (Hg)	ND*	< 0.1	< 0.1
Chromium (Cr)	ND*	< 1	< 1
Tin (Sn)	ND*	< 10	< 10

Arsenic (As2@3)dmium (Cd) Lead (Pb) Mercury (H@hromium (Cr)



Conclusions:

No heavy metal residues detected.

No flammable residues detected.

No chemical residues detected.

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Martin V. Batch# 744 31-July 2017 31-July 2017 01-August 2017

Pesticide Analysis: Our tests looked for residue of

nearly 300 known pesticides finding no evidence

of any over detectable limits.

Endoca Labs tests our products thoroughly. Nearly 300 of the below pesticides concentrations were measured and we are proud to say that all tests measured below our detectable limits. Most tests have a threshold of 0.01 mg/k, while only a handful of tests have a threshold value of <0.05 mg/kg. Not a single test of Endoca products went over detectable threshold limits.

PESTICIDES MEASURED

Acrinathrin Azoxystrobin Biphenhin Bitertanol Biphenyl Bromopropylate Bromuconazole Bupirimate Cadusafos Captafol Captan Chlorphenson Chlorfenapyr Chlorfenvinphos Chlorothalonil Chlorprophame 3,5-Dichloraniline Chlorpyriphos Chlorpyrifos-methyl Chlorthal-dimethyl Cyfluthrin Cypermethrin Cyproconazole Cyprodinil Clomazone o,p-DDE P,P-DDE o,p-DDD P,P-DDD o,p-DDT p,p-DDT Deltamethri Diazinon Diclofop-methyl Dieldrin Dichlobenil Dichlofluanid Dichlorvos Dicloran Dicofol Dicrotophos Diethofencarb Diflubenzuron Dimetachlor Diniconazole Dodemorph Diphenylamine Alpha-Endosulfan Beta-Endosulfan Endosulfan-sulphate Ethion Etofumesate Ethoprophos Ehtoxyquin Etoxazole Etridiazole Etrimphos Famoxadone Fenarimol Fenazaguin Fenchlorphos Fenhexamid Fenihothion Fenpropidin Fenpropimorph Fenvalerate Formothion Fipronil Fipronil-sulfone Fludioxonil Flusilazole Flutriafol Folpet Fuberidazole Furathiocarb Hexaconazole HCB Alpha-HCH Beta-HCH Delta-HCH Heptachlor Heptachlor-epoxidceis Heptachlorepoxidtreans Iprodione Iprovalicarb Lambda- cyhalothrin Lindane Mecarbam Metalaxv Metazachlor Methidathion Metribuzin Mevinphos Myclobutanil Nuarimol Orthophenylphenol Oxadixyl Paclobutrazol Parathion Parathion-methyl Paraoxonmethyl Paraoxon-ethyl Penconazole Pendimethaline Permethrin Phenthoate Phorate Procymidone Profenofos Propiconazole Propyzamide Pyrazophos Pyrethrins Pyridaben Pyrimethanil Pyriproxyfen Quinoxifen Quitozene Pentachloraniline Phosphamidon Pyrifenox Prometryn Propanil Propoxur Proquinazid Prothiofos Simazine Spiroxamine T au-fluvalinate T ebuconazole T ebufenpyrad T ecnazene T efluthrin T erbuthylazine T etraconazole T etradifon T etramethrine T olclofos-methyl T olylfluanid Transfluthrin Triadimephon Triadimenol Trialate Trifloxystrobin Triflumizole Vinclozolin DDT isomersum Heptachlor (heptachloarnd heptachloer poxidsum) Trifluraline Chlorobenzilate 3-Chloraniline Abamectin (AvermectinBla and AvermectinBlb sum) Acetamiprid Aldicarb Aldikarbsulphone Aldicarbsulphoxide Azinphos-ethyl Azinphos-methyl Benalaxyl Benfuracarb Boscalid Buprofezin Carbaryl Carbendazim Carbofuran 3-hydroksicarbofuran Carbosulfan Chloridazon Cymoxanil Clofentezin Clothianidin Demeton-S-methyl Demeton-S-methyslulfoxid Diafenthiuron Difenoconazole Dimethoate Dimethomorph Diuron EPN Epoxiconazole Ethirimol Etofenprox Fenamidone Fenbuconazole Fenbutatinoxid Fenoxycarb Fenpyroximate Fenpropathrin Fensulfothion Fenthion Fenthionsulphone Fenthionsulphoxide Fluazinam Flufenoxuron Fluquinconazole Fonofos Formetanate Fosthiazate Hexythiazox Imazalil Imidacloprid Indoxacarb Isofenphos Methacrifos Isofenphosmethyl Krezoxim-methyl Linuron Lufenuron Malaoxon Malathion Mepanipirim Mepronil Metamitron Metconazole Methamidophos Methiocarb Methiocarbsulphone Methiocarbsulfoxide Methomyl Methoxyfenozide Metobromuron Monocrotophos Monolinuron Omethoate Oxamyl Pencycuron Phenmedipham Phosalone Phosmet Phosmeot xon Phoxim Pymetrozine Piperonylbutoxide Pyraclostrobin Pyridaphenthion Pyridate Pyrifenox Pirimicarb Pirimicarbdesmethyl Pirimiphos-methyl Primisulfuron-methyl Prochloraz Propamocarb Propargite Prothioconazole Prothioconazole-desthio Quinalphos SpinosynA SpinosynD Sulfotep T ebufenozide T eflubenzuron Thiabendazole Thiacloprid Thiamethoxam Thiodicar Thiophanatemethyl Tralkoxydim Triazophos Trichlorfon Triflumuron Triforine Triticonazole Zoxamide Acephate Amitraz Fenamiphos Fenamiphosulphone Fenamiphosulfoxid Nitempiram Fenthionoxonsulphone Fenthionoxonsulfoxid Kumapho Piriphenox Mehibuzine DEET

Our laboratory analysis is standardized after following protocols: LST EN ISO 6579:2003 / AC:2006 / P:2007 LST EN ISO 11290-1:2003 / A1:2004 / P:2005 LST ISO 16649-2:2002 / P:2009 LST ISO 21527-2:2008 Method PLM 486G

Note on Cannabinoid Testing:

All cannabinoids in their acid forms (ending in "-A") are convertible to their non-acid forms via a decarboxylation process (heating). The components lose mass through this process. To find the total theoretical active cannabinoids, one multiplies the acid forms by 87.7%. For example, CBD-A can be converted to active CBD using the formula: CBD-A x 0.877 = CBD. In this

case, the Max CBD for the sample is: Max CBD (%) = (%CBD-A \times 0.877) + %CBD. The same calculation assay is valid for THC-A. This method has been validated according to the principles of the International Conference on Harmonisation.

Chromatographic Analysis:

Analysis of cannabinoids content was performed using Waters 2695 (Milford, MA, USA) separation module equipped with auto injector, sample cooler, vacuum degasser and column heater units. Separation of all cannabinoids was accomplished on YMC PRO C18 (150 x 4 mm I.D., S-3 μ m) RP column coupled with C18 precolumn maintained at 30 °C by a CTO-20AC column oven.

Isocratic elution consisted of acetonitrile:water (4:1) was done in 30 min. The flow rate was maintained at 0.8 ml/min. The cannabinoids CBD, CBG and THC were monitored at 225 and CBDA, CBGA and THC-A were monitored at 300 nm respectively using dual absorbance detector Waters 2487 (Milford, MA, USA). The injection volume of 0.1 mg/ml sample was 10 μ l. Data evaluation was performed using Clarity software.

Quantification of cannabinoids was obtained from linear regression equation of calibration curve of individual reference standard by plotting concentration versus the area ratio.

Sample preparation for HPLC analysis

0.01 g (±.0001) of homogeneous cannabis extract was diluted with 1 ml of methanol (HPLC grade). Solution was sonicated for 5 min and vortexing for 10 sec. Samples before HPLC analysis were centrifuged at 4800 rpm and further diluted with methanol to the final concentration of 1 mg/ml.

Analysis of terpenes was performed using GC-FID system equipped with auto injector. Separation was accomplished on RTX-5 w/Integra-Guard, 30m, 0.25 mm ID column.

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